

Practical Guidance on the Detection of *NTRK* Fusions in Sarcomas: Current Status and Diagnostic Challenges

Orientações Práticas sobre a Detecção de Sarcomas de Fusão *NTRK*: Estado Atual e Desafios Diagnósticos

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ABSTRACT

Sarcomas are a rare and heterogeneous group of mesenchymal malignant tumors and account for approximately 1% of all adult cancers and around 20% of all pediatric solid tumors in Europe. Technology advances have enabled a more accurate and efficient characterization of the molecular mechanisms underlying the pathogenesis of sarcoma subtypes and revealed novel and unexpected therapeutic targets with prognostic/predictive biomarkers, namely the neurotrophic tyrosine receptor kinase (*NTRK*) gene fusion. The *NTRK* fusion assessment has recently become a standard part of management for patients with unresectable locally advanced or metastatic cancers and has been identified in various tumor types. In the more prevalent adult and pediatric sarcomas, *NTRK* fusions are present in 1% and 20%, respectively, and in more than 90% of very rare subsets of tumors. The inhibition of TRK activity with first-generation TRK inhibitors has been found to be effective and well tolerated in adult and pediatric patients, independently of the tumor type. Overall, the therapeutic benefit to those patients compensates for the difficulties of identifying *NTRK* gene fusions. However, the rarity and diagnostic complexity of *NTRK* gene fusions raise several questions and challenges for clinicians. To address these issues, an expert panel of medical and pediatric oncologists, radiologists, surgeons, orthopedists, and pathologists reviewed the recent literature and discussed the current status and challenges, proposing a diagnostic algorithm for identifying *NTRK* fusion sarcomas. The aim of this article is to review the updated information on this issue and to provide the experts' recommendations and practical guidance on the optimal management of patients with soft tissue sarcomas, infantile fibrosarcoma, gastrointestinal stromal tumors, and osteosarcoma.

Keywords: Gene Fusion; Oncogene Proteins, Fusion/genetics; Receptor, trkA/genetics; Sarcoma/genetics

RESUMO

Os sarcomas são um grupo raro e heterogéneo de tumores mesenquimatosos malignos, e constituem um dos principais grupos de cânceros raros na Europa, representando cerca de 1% de todos os cânceros em adultos e cerca de 20% de todos os tumores sólidos pediátricos. Os avanços tecnológicos permitiram uma caracterização mais precisa e eficiente dos mecanismos moleculares subjacentes à patogénese dos subtipos de sarcoma e revelaram novos e inesperados alvos terapêuticos e biomarcadores prognósticos/preditivos, nomeadamente o gene de fusão do recetor tirosina cinase neurotrófico (*NTRK*). A avaliação da fusão de *NTRK* foi incluída, recentemente, na gestão de doentes com cânceros localmente avançados irremediáveis ou metastáticos e foi identificada em vários tipos de tumores de adultos e pediátricos. Nos sarcomas mais prevalentes diagnosticados em adultos e pediátricos, as fusões de *NTRK* estão presentes em 1% e 20%, respetivamente, e em mais de 90% dos subconjuntos de tumores muito raros. A inibição da atividade de TRK com inibidores de primeira geração tem-se mostrado eficaz e bem tolerada em doentes adultos e pediátricos, independentemente do tipo de tumor. Globalmente, o benefício terapêutico para estes doentes compensa as dificuldades em identificar os genes de fusão de *NTRK*, sendo que a raridade e a complexidade diagnóstica dos genes de fusão de *NTRK* levantam várias questões e desafios para os médicos. Para abordar estas questões, um painel

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de oncologistas médicos e pediátricos, radiologistas, cirurgiões, ortopedistas e patologistas reviram a literatura recente e discutiram o estado atual e os desafios, propondo um algoritmo de diagnóstico para identificar sarcomas de fusão de *NTRK*. Este artigo pretende apresentar uma revisão da literatura atual sobre o tema e fornecer as recomendações dos especialistas e orientações práticas para a gestão de doentes com sarcomas de tecidos moles, fibrossarcoma infantil, tumores do estroma gastrointestinal e osteossarcomas.

Palavras-chave: Fusão Génica; Proteínas de Fusão Oncogénica/genética; Receptor trkA/genética; Sarcoma/genética

INTRODUCTION

Sarcomas are a rare and heterogeneous group of mesenchymal malignant tumors, accounting for approximately 1% of all adult cancers and 20% of all pediatric solid tumors.¹⁻⁶ These tumors can occur in virtually any anatomic site, arising in either soft tissue (~80%) or bone (~20%).¹⁻⁵

Complete surgical resection with or without pre-and postoperative therapies is the standard treatment for most localized sarcomas.^{1,7} In locally advanced, metastatic, or recurrent settings, treatment may involve a combination of strategies, including systemic therapy and local approaches.^{1,8} The clinical management of sarcomas remains challenging due to their heterogeneity, aggressive nature, and different responses to current treatment options.^{7,9}

Technological advances have enabled a more accurate and efficient characterization of the molecular mechanisms underlying the pathogenesis of sarcomas and revealed novel therapeutic targets and prognostic/predictive biomarkers.^{1,7,10-12} The discovery of neurotrophic tyrosine receptor kinase (*NTRK*) gene fusions as sarcoma oncogenic drivers led to new personalized therapies for a subset of patients in the form of tropomyosin receptor kinase (TRK) inhibitors, improving clinical outcomes.^{1,12-15}

The *NTRK* fusion assessment has recently become a standard for patients with unresectable locally advanced or metastatic cancers.¹⁶ These fusions can be detected using a variety of methods, including tumor deoxyribonucleic acid (DNA) and ribonucleic acid (RNA) sequencing and plasma cell-free DNA profiling.^{1,13,17}

Although rare in most common tumor types, *NTRK* fusions are recurrent in certain tumors such as secretory carcinoma of the salivary gland, secretory carcinoma of the breast, thyroid cancers, congenital mesoblastic nephroma, pediatric melanoma, infantile gliomas and infantile fibrosarcoma, where they can be present in > 90% of cases.^{1,15,18} In contrast, *NTRK* fusions have been identified in < 1% of other adult and pediatric sarcomas.^{1,15,18}

The rarity and diagnostic complexity of *NTRK* gene fusions raises several questions and challenges for clinicians. To address these issues, an expert panel of Portuguese medical and pediatric oncologists, radiologists, surgeons, orthopedic surgeons, and pathologists reviewed the recent literature and discussed diagnostic challenges of patients with soft tissue sarcomas, infantile fibrosarcoma, gastrointestinal stromal tumors (GIST), and osteosarcoma.

The aim of this article is to present updated information

on this issue and the experts' proposal of a diagnostic algorithm for *NTRK* fusion sarcomas for practical guidance on the optimal management of these patients.

OVERVIEW OF *NTRK* FUSION CANCER

Etiology of *NTRK* Cancers

The *NTRK* gene family includes three members: *NTRK1* (chromosome 1q23.1), *NTRK2* (chromosome 9q21.33), and *NTRK3* (chromosome 15q25.3), that encode transmembrane TRK proteins TRKA, TRKB, and TRKC, respectively.^{1,19} Tropomyosin receptor kinase proteins are expressed in the adult's peripheral and central nervous systems (CNS), and during embryonic development.²⁰

Under normal physiological conditions, TRK proteins bind to neurotrophic family ligands leading to downstream signaling that is critical for the normal development and function of the peripheral and CNS.^{1,19,21} However, *NTRK* genes may undergo chromosomal rearrangements due to intra- and inter-chromosomal translocations of the kinase portion of *NTRK1/2/3* with an unrelated gene.^{20,22} These gene fusions lead to the constitutive activation/expression of chimeric TRK proteins, which have oncogenic properties by driving uncontrolled cell proliferation and tumor growth in a variety of tissues.^{20,22}

Neurotrophic tyrosine receptor kinase fusions were first found in colon carcinoma²³ and later described for the first time in pediatric fibrosarcomas, namely the *ETV6::NTRK3* fusion.²⁴ Currently, there are over 80 different fusion partners identified in a wide range of tumor types.²³⁻²⁵ In rare cancers, the most common detected gene fusion is *ETV6::NTRK3*, while in the more common, the *NTRK* genes can be found with a large number of different partners, with the *NTRK1* gene usually having more fusion partners than *NTRK2* and *NTRK3* genes.^{26,27}

NTRK gene fusions in soft tissue sarcomas

Soft tissue sarcomas comprise a heterogeneous group of cancers with different responses to treatment, which ultimately confer an aggressive behavior, poor prognosis, and a five-year overall survival (OS) rate of 65%.^{28,29} Disease management should be performed by a multidisciplinary team in a sarcoma reference center. Among the underlying causes of soft tissue sarcomas, *NTRK* gene fusions account for only 1% of cases.²⁸⁻³²

Infantile fibrosarcoma

Infantile fibrosarcoma is a rare pediatric tumor that usually occurs in the first year of life. It forms in connective tissue and, in almost 50% of cases, arises in the extremities of the body, followed by the head, neck, and trunk.^{33,34} The tumor often presents a fast-growing period, but it rarely metastasizes. Resection of the tumor with clean margins is the mainstay of treatment, although 48% - 62% are considered unresectable.³⁴ Chemotherapy has been shown to improve OS in infantile fibrosarcoma by improving the ability to remove the tumor. Infantile fibrosarcoma are tumors with a high prevalence of *NTRK* fusions.¹³ *ETV6::NTRK3* fusion is the most common fusion and, together with other variants, is present in about 90% of infantile fibrosarcomas.^{24,35,36} This implies a new therapeutic target for these patients.

Gastrointestinal stromal tumors

Gastrointestinal stromal tumors are rare tumors characterized by usually small gastroesophageal or duodenal nodules with a progression risk associated with tumor size and mitotic index.^{37,38} In adults, GIST typically occurs in patients aged 60 - 65 and frequently harbor *KIT* or *PDGFRA* mutations.^{38,39} Pediatric GIST is mainly characterized by loss of function mutations in succinate dehydrogenase (SDH) genes, encoding the subunits of the SDH enzyme.⁴⁰ Regarding *NTRK* gene fusion frequency, screening of 24 GIST lacking *KIT* or *PDGFRA* mutations showed one *NTRK* fusion-positive tumor (4.2%).⁴¹ However, another study of targeted sequencing data from 738 GIST did not find cases with *NTRK* rearrangements. More comprehensive large-scale studies are needed to confirm *NTRK* fusion incidence in GIST.

Osteosarcoma

Osteosarcoma has an overall incidence of 0.3 cases per 100 000/year and occurs most frequently around the knee in adolescents and in the craniofacial bones in adults.⁴² High-grade osteosarcoma patients usually develop metastases in the lungs and distant bones.⁴³ *NTRK* fusions appear to be rare in bone sarcomas, as suggested by a study comprising 354 bone tumors that did not find *NTRK* gene fusion after immunohistochemistry screening.^{44,45}

ASSESSMENT OF *NTRK* FUSIONS IN SARCOMAS

Technologies for testing *NTRK* fusions

New *NTRK* gene fusions are being discovered regularly, resulting from the emergence of new screening methodologies.³⁰ The most common technologies to detect, directly or indirectly, *NTRK* fusions in tumor tissues are immunohistochemistry (IHC), reverse transcriptase polymerase chain reaction (RT-PCR), fluorescence *in situ* hybridization (FISH), next-generation sequencing (NGS) of DNA and/or

RNA, and NanoString nCounter technique.

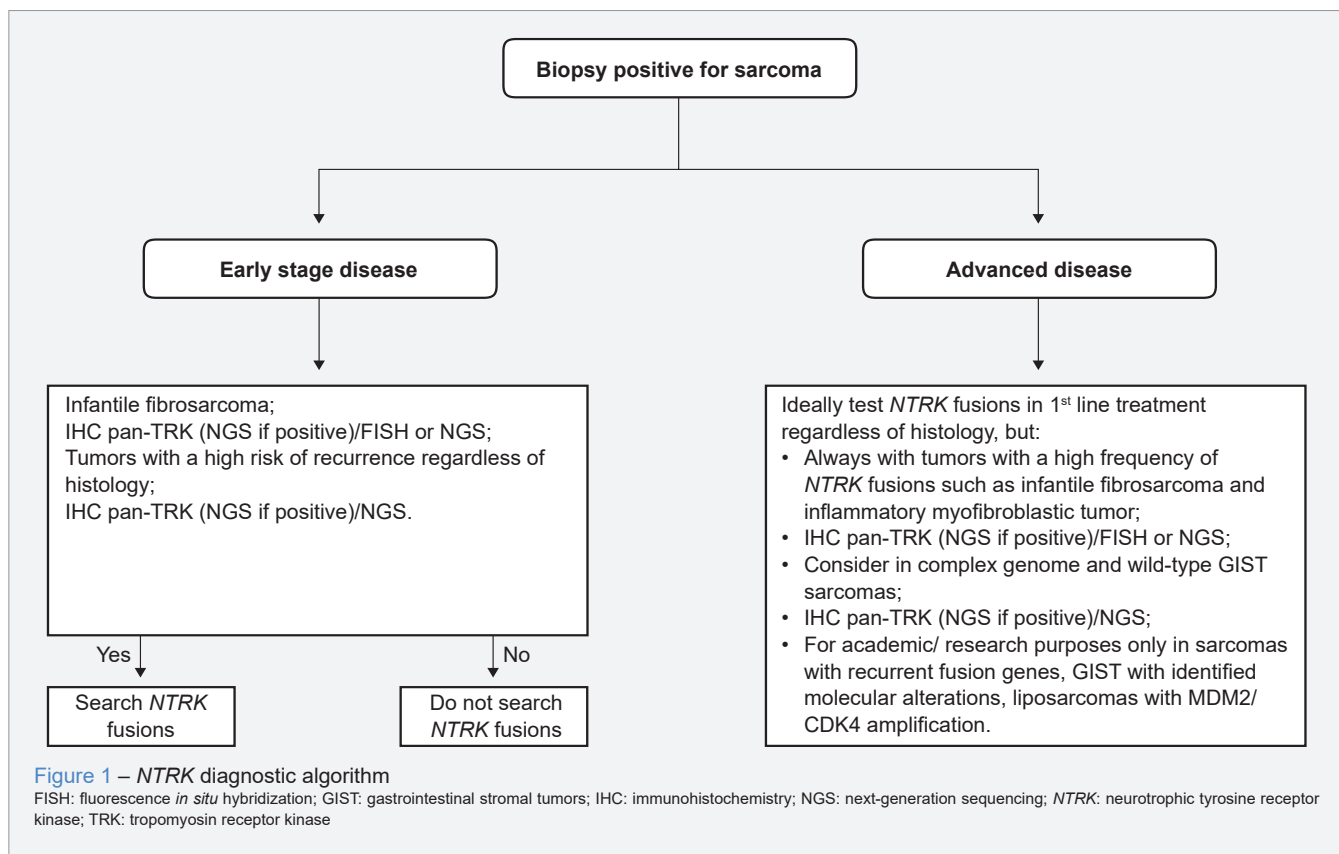
Immunohistochemistry

Immunohistochemistry is a valuable screening tool in clinical environments with limited access to NGS platforms. Regarding other methods, IHC has several benefits, namely time- and tissue-efficiency with an overall good cost-effectiveness.⁴⁶ This technique is highly sensitive for detecting *NTRK1/2* fusions but sub-optimal for *NTRK3* fusions (sensitivity < 79%).¹⁶ The anti-TRKA and pan-TRK antibodies can be used to spot elevated TRK expression compared to the low TRK levels observed in control cells.^{15,16,46} These antibodies enable the detection of the gene fusions at the protein level, allowing to distinguish between expressing (detectable) and non-expressing (non-detectable) *NTRK* fusions. The staining pattern can be correlated with the subcellular location of the *NTRK* fusion partner. However, TRKA/B/C proteins are physiologically expressed in some healthy cells, like neural and muscle tissue, making it difficult to evaluate the presence of *NTRK* fusions in tumors derived from or involving such organ systems. Additionally, sample preparation can lead to false negatives. Hence, internal and external controls, such as endothelial cells and positive cell lines, are highly recommended. The absence of standard criteria for immunohistochemistry evaluation complicates the interpretation of IHC data; thus, positive results should be followed with a molecular method to further confirm the presence of *NTRK* fusion.¹⁶

Although international guidelines recommend confirmation of positive TRK IHC with a targeted RNA analysis, up-front testing with a targeted RNA analysis should be preferred in some scenarios since there is limited evidence available regarding the use of IHC in detecting *NTRK* gene alterations in routine practice.^{1,15,47,48}

Reverse transcriptase polymerase chain reaction

Reverse transcriptase polymerase chain reaction is a well-established technique to measure the expression of fusion transcripts implicated in a wide variety of sarcomas.⁴⁹⁻⁵¹ This method employs a 3' primer annealing to an *NTRK* kinase domain and a 5' primer annealing to a fusion partner, flanking the fusion region.⁵² In the presence of the targeted region, the aid of fluorescent signaling probes at each PCR cycle allows detecting the DNA amplification with high sensitivity and specificity.⁵² Reverse transcriptase polymerase chain reaction can also be used for quantitative reporting of tumor burden or post-treatment monitoring. One disadvantage of RT-PCR is the need to design a set of primers for each gene fusion transcript that, together with an increasing number of 5' fusion partner genes, reduces the applicability of a multiplex RT-PCR assay.⁵³ Moreover, it is restricted to known fusion partners, which can lead to false negatives in



benefit of patient's healthcare.

CLINICAL MANAGEMENT OF *NTRK*-FUSED SARCOMAS

Therapies for patients with TRK fusion sarcoma

Several small molecules, grouped into multi-kinase inhibitors or more-selective TRK inhibitors, with different levels of affinity to the TRK domain, are currently in clinical trials (CTs) and some are already approved.⁶⁴ Many of them have demonstrated efficacy in *NTRK* fusion-positive solid tumors.³¹ The multi-kinase inhibitor group includes entrectinib, crizotinib, cabozantinib, lestaurtinib, ponatinib, nintedanib, merestinib, MGCD516, PLX7486, DS-6051b, and TSR-011.³¹ The most specific TRK inhibitor is larotrectinib, the first FDA-approved TRK inhibitor. Larotrectinib and entrectinib are now the first-generation of TRK inhibitors approved for adult and pediatric patients who have a solid tumor with a *NTRK* fusion and no acquired-resistance mutations, which is metastatic or unresectable and a relapse prior to therapy, or without satisfactory alternative treatment options.

FIRST-GENERATION TRK INHIBITORS

Larotrectinib

Larotrectinib is a highly effective and highly selective pan-TRK inhibitor,^{2,65} with a binding affinity capacity of more

than 100-fold when compared with a panel of several kinases.⁶⁶ It has demonstrated a robust tumor-agnostic effect in various sarcomas, including osteosarcoma, dedifferentiated chondrosarcoma, GIST, infantile fibrosarcoma and other soft tissue sarcomas (adult fibrosarcoma, inflammatory myofibroblastic tumor, infantile myofibromatosis, lipofibromatosis, malignant peripheral nerve sheath tumor, myopericytoma, spindle cell sarcoma, high-grade endometrial stromal tumor, and synovial sarcoma).^{1,65}

On the clinical setting, larotrectinib has demonstrated a high efficacy profile in a pooled analysis of the first 55 consecutively enrolled patients of three phase I/II clinical trials (CTs) in adult and pediatric TRK fusion-positive cancers, regardless of patient age or tumor type.⁵⁷ The overall response rate (ORR) was 75%, the median time of response was 1.8 months and, after one year, 71% of responses were ongoing, with 55% of all patients remaining progression-free.⁵⁶ In a recent pooled analysis of the same CTs, including 159 patients with TRK fusion-positive cancer aged from < one month to 84 years and treated with larotrectinib, an objective response of 79% (95% CI 72 - 85) was reported, with 16% having complete responses.⁶⁸

Larotrectinib is available in oral, liquid, or capsule formulations with similar pharmacokinetics, which allows proper administration in infants and children.⁶⁹ Additionally,

promising results in inhibiting most on-target *NTRK* resistance mutations, and currently, is in phase I/II CTs to establish safety, dosing, and clinical efficacy.⁹⁴

ISSUES AND QUESTIONS ON *NTRK* FUSION SARCOMAS

Testing difficulties

Each molecular diagnosis technique has advantages and disadvantages. Testing decisions should ultimately be made based on the type of tumor and the resources available, including the quality and quantity of biopsy material and equipment accessibility.¹⁶

Although costly, RNA-based NGS is the gold standard to test *NTRK* gene fusions in sarcomas. IHC is well accepted as a pre-screening tool, but it gives a high rate of false-negative staining in the case of *NTRK3* fusions. RT-PCR and FISH are highly sensitive techniques; however, the former only detects previously known *NTRK* gene fusions, while the latter may not detect some rearrangements derived from small genomic deletions.^{16,18,96}

These technologies are optimized to work in formalin-fixed paraffin-embedded sample tissue, and it is important to have an image-guided biopsy to collect the material.^{30,51,55}

More recently, some technologies have been developed to take advantage of liquid biopsies, from which circulating tumor cells and circulating cell free tumor DNA/RNA can be harvested.^{96,97} Circulating tumor DNA represents a non-invasive approach that allows monitoring tumor recurrence or progression throughout treatment. However, the sensitivity level of this method will vary with the cell shedding capacity of the tumor and, consequently, with the amount of material for detection in circulation.⁹⁷

The FISH and IHC methods have already been optimized to directly detect gene rearrangements in filtration-enriched circulating tumor cells from NSCLC.⁹⁶ Still, validation from other groups is needed before clinical implementation.

Genetic variability and mutations

A variety of *NTRK* alterations, other than fusions, have been identified in 14% of several tumor types, including point mutations, amplifications, deletions, and splice variants.⁹⁸ Data showing the response of tumors with non-fusion *NTRK* alterations treated with TRK inhibitors is still limited. A case-report presented one patient with an *NTRK* amplification that exhibited a partial response of short duration; however, none of the tumors with *NTRK* point mutations responded to treatment.⁶⁹ Another described a patient with a metastatic esophageal squamous cell carcinoma harboring an *NTRK1* amplification treated with larotrectinib. Initially, the patient showed a partial response of the primary and metastatic tumors, but 3.5 months later, the disease progressed.⁹⁹

CONCLUSION

Since *NTRK* fusions are present in 1% to 20% of the more prevalent adult and pediatric sarcomas, and more than 90% of very rare subsets of tumors, patients eligible for TRK inhibitors are a minority within the overall number of cases of patients with sarcoma.^{1,18} Nevertheless, the inhibition of TRK activity with first-generation of TRK inhibitors is effective and well tolerated in adult and pediatric patients, independently of the tumor type.^{69,70,87}

The therapeutic benefit to those patients compensates for the difficulties of identifying *NTRK* gene fusions. Accordingly, pathologists play a critical role in the diagnosis and assessment of patients with cancer. Several clinical guidelines and *NTRK* gene fusion testing recommendations have been developed to help identify *NTRK* fusion-positive cancers.^{63,100} Following these diagnostic algorithms, pathologists should consider the optimal use of tumor tissue and testing prioritization when tumor tissue is limited, such as small biopsies and cytological samples.

In this manuscript, we have reviewed the etiology of *NTRK* cancers and gene fusions in soft tissue sarcomas, namely infantile fibrosarcoma, GIST, and osteosarcoma, and the therapies for patients with TRK fusion sarcoma, including first- and next-generation TRK inhibitors. We reviewed the technologies for testing *NTRK* fusions and discussed the diagnostic challenges. Aiming at optimizing clinical management of these patients we propose a diagnostic algorithm for identifying *NTRK* fusion sarcomas (Fig. 1).

In Portugal, evidence is limited due to regulatory issues. Despite the most recent data and the consensus among the participants in this working group, there is no public coverage in Portugal for these medicines, limiting patients' access to therapeutics. Real-world evidence studies will be essential to demonstrate the improvement in survival with QoL for sarcoma patients with *NTRK* fusion.

AUTHOR CONTRIBUTIONS

All authors participated in the consensus-elaboration meeting and contributed to the diagnostic algorithm elaboration, paper revision and validation.

COMPETING INTERESTS

IF received research funding from PharmaMar and Roche.

AF received honoraria for presentations from Novartis, AstraZeneca and Gilead; support for attending meetings and travel from Pfizer and Gilead; participated on an advisory board from AstraZeneca, Daichi and Lilly; has a leadership role in the Portuguese Oncologic Study Group.

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HV received medical writing support from Bayer; has a leadership role in Grupo Português de Estudos em Sarcomas.

All other authors have declared that no competing interests exist.

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REFERENCES

- Demetri GD, Antonescu CR, Bjerkeheggen B, Bovée JV, Boye K, Chacón M, et al. Diagnosis and management of tropomyosin receptor kinase (TRK) fusion sarcomas: expert recommendations from the World Sarcoma Network. *Ann Oncol*. 2020;31:1506-17.
- Stiller CA, Trama A, Serraino D, Rossi S, Navarro C, Chirlaque MD, et al. Descriptive epidemiology of sarcomas in Europe: report from the RARECARE project. *Eur J Cancer*. 2013;49:684-95.
- Agnoletto C, Caruso C, Garofalo C. Heterogeneous circulating tumor cells in sarcoma: implication for clinical practice. *Cancers*. 2021;13:2189.
- McConnell L, Houghton O, Stewart P, Gazdova J, Srivastava S, Kim C, et al. A novel next generation sequencing approach to improve sarcoma diagnosis. *Mod Pathol*. 2020;33:1350-9.
- Szurian K, Kashofer K, Liegl-Atzwanger B. Role of next-generation sequencing as a diagnostic tool for the evaluation of bone and soft-tissue tumors. *Pathobiology*. 2018;84:323-38.
- Siegel RL, Miller KD, Fuchs HE, Jemal A. Cancer Statistics, 2021. *CA Cancer J Clin*. 2021;71:7-33.
- Bleloch JS, Ballim RD, Kimani S, Parkes J, Panieri E, Willmer T, et al. Managing sarcoma: where have we come from and where are we going? *Ther Adv Med Oncol*. 2017;9:637-59.
- Dangoor A, Seddon B, Gerrand C, Grimer R, Whelan J, Judson I. UK guidelines for the management of soft tissue sarcomas. *Clin Sarcoma Res*. 2016;6:1-26.
- Damerell V, Pepper MS, Prince S. Molecular mechanisms underpinning sarcomas and implications for current and future therapy. *Signal Transduct Target Ther*. 2021;6:246.
- Grünewald TG, Alonso M, Avnet S, Banito A, Burdach S, Cidre-Aranaz F, et al. Sarcoma treatment in the era of molecular medicine. *EMBO Mol Med*. 2020;12:1-33.
- Gómez J, Tsagozis P. Multidisciplinary treatment of soft tissue sarcomas: an update. *World J Clin Oncol*. 2020;11:180-9.
- Xu L, Xie X, Shi X, Zhang P, Liu A, Wang J, et al. Potential application of genomic profiling for the diagnosis and treatment of patients with sarcoma. *Oncology Lett*. 2021;21:1-12.
- Cocco E, Scaltriti M, Drilon A. NTRK fusion-positive cancers and TRK inhibitor therapy. *Nat Rev Clin Oncol*. 2018;15:731-47.
- Kheder ES, Hong DS. Emerging targeted therapy for tumors with NTRK fusion proteins. *Clin Cancer Res*. 2018;24:5807-14.
- Brčić I, Godschachner TM, Bergovec M, Igrec J, Till H, Lackner H, et al. Broadening the spectrum of NTRK rearranged mesenchymal tumors and usefulness of pan-TRK immunohistochemistry for identification of NTRK fusions. *Mod Pathol*. 2021;34:396-407.
- Solomon JP, Linkov I, Rosado A, Mullaney K, Rosen EY, Frosina D, et al. NTRK fusion detection across multiple assays and 33,997 cases: diagnostic implications and pitfalls. *Mod Pathol*. 2020;33:38-46.
- Miettinen M, Felisiak-Golabek A, Luiña Contreras A, Glod J, Kaplan RN, Killian JK, et al. New fusion sarcomas: histopathology and clinical significance of selected entities. *Hum Pathol*. 2019;86:57-65.
- Siozopoulou V, Smits E, De Winne K, Marcq E, Pauwels P. NTRK fusions in sarcomas: diagnostic challenges and clinical aspects. *Diagnostics*. 2021;11:478.
- Hechtman JF. NTRK insights: best practices for pathologists. *Mod Pathol*. 2022;35:298-305.
- Simmons C, Deyell RJ, MacNeill AJ, Vera-Badillo FE, Smrke A, Abdul Razak AR, et al. Canadian consensus on TRK-inhibitor therapy for NTRK fusion-positive sarcoma. *Int J Cancer*. 2021;149:1691-704.
- Lassen U. How I treat NTRK gene fusion-positive cancers. *ESMO Open*. 2019;4:S612.
- Vaishnavi A, Le AT, Doebele RC. TRKING down an old oncogene in a new era of targeted therapy. *Cancer Discov*. 2015;5:25-34.
- Martin-Zanca D, Hughes SH, Barbacid M. A human oncogene formed by the fusion of truncated tropomyosin and protein tyrosine kinase sequences. *Nature*. 1986;319:743-8.
- Knezevich SR, McFadden DE, Tao W, Lim JF, Sorensen PH. A novel ETV6-NTRK3 gene fusion in congenital fibrosarcoma. *Nat Genet*. 1998;18:184-7.
- Lange AM, Lo HW. Inhibiting TRK proteins in clinical cancer therapy. *Cancers*. 2018;10:105.
- Kummar S, Lassen UN. TRK Inhibition: a new tumor-agnostic treatment strategy. *Target Oncol*. 2018;13:545-56.
- Amatu A, Sartore-Bianchi A, Siena S. NTRK gene fusions as novel targets of cancer therapy across multiple tumour types. *ESMO open*. 2016;1:e000023.
- Ardakani AH, Ware H, Woollard A, Gikas P. Soft tissue sarcoma: recognizing a rare disease. *Cleve Clin J Med*. 2022;89:73-80.
- Gronchi A, Miah AB, Dei Tos AP, Abecassis N, Bajpai J, Bauer S, et al. Soft tissue and visceral sarcomas: ESMO–EURACAN–GENTURIS Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol*. 2021;32:1348-65.
- Westphalen CB, Krebs MG, Le Tourneau C, Sokol ES, Maund SL, Wilson TR, et al. Genomic context of NTRK1/2/3 fusion-positive tumours from a large real-world population. *NPJ Precis Oncol*. 2021;5:1-9.
- Assi T, Rassy E, Nassereddine H, Farhat F, Karak FE, Kattan J, et al. TRK inhibition in soft tissue sarcomas: a comprehensive review. *Semin Oncol*. 2020;47:73-84.
- Zhao X, Kotch C, Fox E, Surrey LF, Wertheim GB, Baloch ZW, et al. NTRK fusions identified in pediatric tumors: the frequency, fusion partners, and clinical outcome. *JCO Precis Oncol*. 2021;1:PO.20.00250.
- Orbach D, Brennan B, De Paoli A, Gallego S, Mudry P, Francotte N, et al. Conservative strategy in infantile fibrosarcoma is possible: the European paediatric soft tissue sarcoma study group experience. *Eur J Cancer*. 2016;57:1-9.
- Orbach D, Sparber-Sauer M, Laetsch TW, Minard-Colin V, Bielack SS, Casanova M, et al. Spotlight on the treatment of infantile fibrosarcoma in the era of neurotrophic tropomyosin receptor kinase inhibitors: International consensus and remaining controversies. *Eur J Cancer*. 2020;137:183-92.
- Sheng WQ, Hisaoka M, Okamoto S, Tanaka A, Meis-Kindblom JM, Kindblom LG, et al. Congenital-infantile fibrosarcoma. A clinicopathologic study of 10 cases and molecular detection of the ETV6-NTRK3 fusion transcripts using paraffin-embedded tissues. *Am J Clin Pathol*. 2001;115:348-55.
- Albert CM, Davis JL, Federman N, Casanova M, Laetsch TW. TRK fusion cancers in children: a clinical review and recommendations for screening. *J Clin Oncol*. 2019;37:513-24.
- Casali PG, Abecassis N, Bauer S, Biagini R, Bielack S, Bonvalot S, et al. Gastrointestinal stromal tumours: ESMO-EURACAN clinical practice guidelines for diagnosis, treatment and follow-up. *Ann Oncol*. 2018;29:iv68-78.
- Dudzisz-Śledź M, Bylina E, Teterycz P, Rutkowski P. Treatment of metastatic gastrointestinal stromal tumors (GIST): a focus on older patients. *Drugs Aging*. 2021;38:375-96.
- Lasota J, Miettinen M. KIT and PDGFRA mutations in gastrointestinal stromal tumors (GISTs). *Semin Diagn Pathol*. 2006;23:91-102.
- Quiroz HJ, Willobe BA, Sussman MS, Fox BR, Thorson CM, Sola JE, et al. Pediatric gastrointestinal stromal tumors—a review of diagnostic

- modalities. *Transl Gastroenterol Hepatol*. 2018;3:1-8.
41. Shi E, Chmielecki J, Tang CM, Wang K, Heinrich MC, Kang G, et al. FGFR1 and NTRK3 actionable alterations in "wild-type" gastrointestinal stromal tumors. *J Transl Med*. 2016;14:1-11.
 42. Strauss SJ, Frezza AM, Abecassis N, Bajpai J, Bauer S, Biagini R, et al. Bone sarcomas: ESMO-EURACAN-GENTURIS-ERN paedcan clinical practice guideline for diagnosis, treatment and follow-up. *Ann Oncol*. 2021;32:1520-36.
 43. Odri GA, Tchicaya-Bouanga J, Yoon DJ, Modrowski D. Metastatic progression of osteosarcomas: a review of current knowledge of environmental versus oncogenetic drivers. *Cancers*. 2022;14:1-16.
 44. Ameline B, Saba KH, Kovac M, Magnusson L, Witt O, Bielack S, et al. NTRK fusions in osteosarcoma are rare and non-functional events. *J Pathol Clin Res*. 2020;6:107-12.
 45. Lam SW, Briaire-de-Brujin IH, van Wezel T, Cleven AH, Hogendoorn PC, Cleton-Jansen AM, et al. NTRK fusions are extremely rare in bone tumours. *Histopathology*. 2021;79:880-5.
 46. Hechtman JF, Benayed R, Hyman DM, Drilon A, Zehir A, Frosina D, et al. Pan-Trk immunohistochemistry is an efficient and reliable screen for the detection of NTRK fusions. *Am J Surg Pathol*. 2017;41:1547-51.
 47. Marchiò C, Scaltriti M, Ladanyi M, lafrate AJ, Bibeau F, Dietel M, et al. ESMO recommendations on the standard methods to detect NTRK fusions in daily practice and clinical research. *Ann Oncol*. 2019;30:1417-27.
 48. Karakas C, Giampoli EJ, Love T, Hicks DG, Velez MJ. Validation and interpretation of Pan-TRK immunohistochemistry: a practical approach and challenges with interpretation. *Diagn Pathol*. 2024;19:10.
 49. Agaram NP, Zhang L, Sung YS, Chen CL, Chung CT, Antonescu CR, et al. Recurrent NTRK1 gene fusions define a novel subset of locally aggressive lipofibromatosis-like neural tumors. *Am J Surg Pathol*. 2016;40:1407-16.
 50. Tvrdik D, Povýsil C, Svatosová J, Dundr P. Molecular diagnosis of synovial sarcoma: RT-PCR detection of SYT-SSX1/2 fusion transcripts in paraffin-embedded tissue. *Med Sci Monit*. 2005;11:MT1-7.
 51. Ueno-Yokohata H, Okita H, Nakasato K, Kiyotani C, Kato M, Matsumoto K, et al. Establishment of multiplex RT-PCR to detect fusion genes for the diagnosis of Ewing sarcoma. *Diagn Pathol*. 2021;16:1-10.
 52. Bourgeois JM, Knezevich SR, Mathers JA, Sorensen PH. Molecular detection of the ETV6-NTRK3 gene fusion differentiates congenital fibrosarcoma from other childhood spindle cell tumors. *Am J Surg Pathol*. 2000;24:937-46.
 53. Beadling C, Wald AI, Warrick A, Neff TL, Zhong S, Nikiforov YE, et al. A multiplexed amplicon approach for detecting gene fusions by next-generation sequencing. *J Mol Diagn*. 2016;18:165-75.
 54. Chrzanowska NM, Kowalewski J, Lewandowska MA. Use of fluorescence in situ hybridization (FISH) in diagnosis and tailored therapies in solid tumors. *Molecules*. 2020;25:1-21.
 55. Kerr KM, López-Ríos F. Precision medicine in NSCLC and pathology: how does ALK fit in the pathway? *Ann Oncol*. 2016;27:iii16-iii24.
 56. Drilon A, Laetsch TW, Kummar S, DuBois SG, Lassen UN, Demetri GD, et al. Efficacy of larotrectinib in TRK fusion-positive cancers in adults and children. *N Engl J Med*. 2018;378:731-9.
 57. Gao X, Sholl LM, Nishino M, Heng JC, Jänne PA, Oxnard GR. Clinical implications of variant ALK FISH rearrangement patterns. *J Thor Oncol*. 2015;10:1648-52.
 58. Hsiao SJ, Zehir A, Sireci AN, Aisner DL. Detection of tumor NTRK gene fusions to identify patients who may benefit from tyrosine kinase (TRK) inhibitor therapy. *J Mol Diagn*. 2019;21:553-71.
 59. Pfarr N, Kirchner M, Lehmann U, Leichsenring J, Merkelbach-Bruse S, Glade J, et al. Testing NTRK testing: wet-lab and in silico comparison of RNA-based targeted sequencing assays. *Genes Chromosomes Cancer*. 2020;59:178-88.
 60. Salmon CR, Silvério KG, Giorgetti AP, Sallum EA, Casati MZ, Nociti FH. Gene expression analysis in microdissected samples from decalcified tissues. *Diagn Mol Pathol*. 2012;21:120-6.
 61. Goytain A, Ng T. NanoString ncounter technology: high-throughput RNA validation. *Methods Mol Biol*. 2020;2079:125-39.
 62. de Oliveira Cavagna R, de Andrade ES, Tadin Reis M, de Paula FE, Noriz Berardinelli G, Bonatelli M, et al. Detection of NTRK fusions by RNA-based nCounter is a feasible diagnostic methodology in a real-world scenario for non-small cell lung cancer assessment. *Sci Rep*. 2023;13:21168.
 63. Chilimoniuk J, Erol A, Rödiger S, Burdukiewicz M. Challenges and opportunities in processing NanoString nCounter data. *Comput Struct Biotechnol J*. 2024;23:1951-8.
 64. Kummar S, Italiano A, Brose MS, Carlson JJ, Sullivan SD, Lassen U, et al. Diagnosis and management of TRK fusion cancer. *Am J Manag Care*. 2022;28:S15-25.
 65. Doebele RC, Davis LE, Vaishnavi A, Le AT, Estrada-Bernal A, Keyser S, et al. An oncogenic NTRK fusion in a patient with soft-tissue sarcoma with response to the tropomyosin-related kinase inhibitor LOXO-101. *Cancer Discov*. 2015;5:1049-57.
 66. Ghilardi JR, Freeman KT, Jimenez-Andrade JM, Mantyh WG, Bloom AP, Kuskowski MA, et al. Administration of a tropomyosin receptor kinase inhibitor attenuates sarcoma-induced nerve sprouting, neuroma formation and bone cancer pain. *Mol Pain*. 2010;6:87.
 67. Kummar S, Shen L, Hong DS, McDermott R, Keedy VL, Casanova M, et al. Larotrectinib efficacy and safety in a patient with tropomyosin receptor kinase fusion sarcomas. *Cancer*. 2023;129:3772-82.
 68. Hong DS, DuBois SG, Kummar S, Farago AF, Albert CM, Rohrberg KS, et al. Larotrectinib in patients with TRK fusion-positive solid tumours: a pooled analysis of three phase 1/2 clinical trials. *Lancet Oncol*. 2020;21:531-40.
 69. Laetsch TW, DuBois SG, Mascarenhas L, Turpin B, Federman N, Albert CM, et al. Larotrectinib for paediatric solid tumours harbouring NTRK gene fusions: phase 1 results from a multicentre, open-label, phase 1/2 study. *Lancet Oncol*. 2018;19:705-14.
 70. Hong DS, Bauer TM, Lee JJ, Dowlati A, Brose MS, Farago AF, et al. Larotrectinib in adult patients with solid tumours: a multi-centre, open-label, phase I dose-escalation study. *Ann Oncol*. 2019;30:325-31.
 71. Yang AT, Laetsch TW. Safety of current treatment options for NTRK fusion-positive cancers. *Expert Opin Drug Saf*. 2023;22:1073-89.
 72. McDermott R, van Tilburg CM, Farago AF, Kummar S, Tan DS, Albert CM, et al. 1955P survival benefits of larotrectinib in an integrated dataset of patients with TRK fusion cancer. *Ann Oncol*. 2020;31:S1101-2.
 73. Lin JJ, Kummar S, Tan DS, Lassen UN, Leyvraz S, Liu Y, et al. Long-term efficacy and safety of larotrectinib in patients with TRK fusion-positive lung cancer. *J Clin Oncol*. 2021;39:S9109.
 74. Kummar S, Van Tilburg CM, Albert CM, Berlin J, Farago AF, McDermott RS, et al. Quality of life of adults and children with TRK fusion cancer treated with larotrectinib compared to the general population. *J Clin Oncol*. 2020;38:S3614.
 75. Ziegler DS, Wong M, Mayoh C, Kumar A, Tsoli M, Mould E, et al. Brief report: potent clinical and radiological response to larotrectinib in TRK fusion-driven high-grade glioma. *Br J Cancer*. 2018;119:693-6.
 76. Ziegler DS, Doz F, Geoerger B, Dubois S, Grilley-Olson JE, van Tilburg C, et al. Activity of larotrectinib in TRK fusion cancer patients with primary central nervous system tumours. *Ann Oncol*. 2019;30:ix124.
 77. Drilon AE, DuBois SG, Farago AF, Geoerger B, Grilley-Olson JE, Hong DS, et al. Activity of larotrectinib in TRK fusion cancer patients with brain metastases or primary central nervous system tumors. *J Clin Oncol*. 2019;37:S2006.
 78. Doz F, van Tilburg CM, Geoerger B, Højgaard M, Øra I, Boni V, et al. Efficacy and safety of larotrectinib in TRK fusion-positive primary central nervous system tumors. *Neuro Oncol*. 2022;24:997-1007.
 79. DuBois SG, Laetsch TW, Federman N, Turpin BK, Albert CM, Nagasubramanian R, et al. The use of neoadjuvant larotrectinib in the management of children with locally advanced TRK fusion sarcomas. *Cancer*. 2018;124:4241-7.
 80. Anderson D, Ciomei M, Banfi P, Cribioli S, Ardini E, Galvani A, et al. Inhibition of Trk-driven tumors by the pan-Trk inhibitor RXDX-101. *Eur J Cancer*. 2014;50:101.
 81. Ardini E, Menichincheri M, Banfi P, Bosotti R, De Ponti C, Pulci R, et al. Entrectinib, a Pan-TRK, ROS1, and ALK inhibitor with activity in multiple molecularly defined cancer indications. *Mol Cancer Ther*. 2016;15:628-39.
 82. Farago AF, Le LP, Zheng Z, Muzikansky A, Drilon A, Patel M, et al. Durable clinical response to entrectinib in NTRK1-rearranged non-small

- cell lung cancer. *J Thorac Oncol.* 2015;10:1670-4.
83. Demetri GD, Paz-Ares L, Farago AF, Liu SV, Chawla SP, Tosi D, et al. Efficacy and safety of entrectinib in patients with NTRK fusion-positive (NTRK-fp) tumors: pooled analysis of STARTRK-2, STARTRK-1 and ALKA-372-001. *Ann Oncol.* 2018;29:viii713.
 84. Demetri GD, De Braud F, Dilon A, Siena S, Patel MR, Cho BC, et al. Updated integrated analysis of the efficacy and safety of entrectinib in patients with NTRK fusion-positive solid tumors. *Clin Cancer Res.* 2022;28:1302-12.
 85. Doebele RC, Dilon A, Paz-Ares L, Siena S, Shaw AT, Farago F, et al. Efficacy and safety of entrectinib in patients with advanced or metastatic NTRK fusion-positive solid tumours: integrated analysis of three phase 1-2 trials Robert. *Lancet Oncol.* 2020;21:271-82.
 86. Desai AV, Robinson GW, Basu EM, Foster J, Gauvain K, Sabnis A, et al. Updated entrectinib data in children and adolescents with recurrent or refractory solid tumors, including primary CNS tumors. *J Clin Oncol.* 2020;38:S107.
 87. Perreault S, Chami R, Deyell RJ, Demellawy D El, Ellezam B, Jabado N, et al. Canadian consensus for biomarker testing and treatment of TRK fusion cancer in pediatric patients. *Curr Oncol.* 2021;28:346-66.
 88. Rangaraju S, Li G, Christiansen J, Hornby Z, Multani P, Esquibel V, et al. TRTH-10. pediatric phase 1/1b study of entrectinib in patients with primary brain tumors, neuroblastoma, and NTRK, ROS1, or ALK fusions. *Neuro Oncol.* 2017;19:Siv53.
 89. Paz-Ares L, Barlesi F, Siena S, Ahn MJ, Dilon A, Conley A, et al. Patient-reported outcomes from STARTRK-2: a global phase II basket study of entrectinib for ROS1 fusion-positive non-small-cell lung cancer and NTRK fusion-positive solid tumours. *ESMO Open.* 2021;6:100113.
 90. Cocco E, Schram AM, Kulick A, Misale S, Won HH, Yaeger R, et al. Resistance to TRK inhibition mediated by convergent MAPK pathway activation. *Nat Med.* 2019;25:1422-7.
 91. Russo M, Misale S, Wei G, Siravegna G, Crisafulli G, Lazzari L, et al. Acquired resistance to the TRK inhibitor entrectinib in colorectal cancer. *Cancer Discov.* 2016;6:36-44.
 92. Dilon A, Ou SHI, Cho BC, Kim DW, Lee J, Lin JJ, et al. Repotrectinib (Tpx-0005) is a next-generation ROS1/TRK/ALK inhibitor that potently inhibits ROS1/TRK/ALK solvent-front mutations. *Cancer Discov.* 2018;8:1227-36.
 93. Dilon A, Nagasubramanian R, Blake JF, Ku N, Ebata K, Smith S, et al. A next-generation TRK kinase inhibitor overcomes acquired resistance to prior trk kinase inhibition in patients with TRK fusion-positive solid tumors. *Cancer Discov.* 2017;7:963-72.
 94. Murray BW, Rogers E, Zhai D, Deng W, Chen X, Sprengeler PA, et al. Molecular characteristics of repotrectinib that enable potent inhibition of TRK fusion proteins and resistant mutations. *Mol Cancer Ther.* 2021;20:2446-56.
 95. Weiss LM, Funari VA. NTRK fusions and Trk proteins: what are they and how to test for them. *Hum Path.* 2021;112:59-69.
 96. Catelain C, Pailler E, Oulhen M, Faugeroux V, Pommier AL, Farace F. Detection of gene rearrangements in circulating tumor cells: examples of ALK-, ROS1-, RET-rearrangements in non-small-cell lung cancer and ERG-rearrangements in prostate cancer. *Adv Exp Med Biol.* 2017;994:169-79.
 97. Tsoi KM, Wunder JS, Gokgoz N, Darville-O'Quinn P, Prochazka P, Malekoltajari A, et al. Detection and utility of cell-free and circulating tumour DNA in bone and soft-tissue sarcomas. *Bone Joint Res.* 2021;10:602-10.
 98. Okamura R, Boichard A, Kato S, Sicklick JK, Bazhenova L, Kurzrock R. Analysis of NTRK alterations in pan-cancer adult and pediatric malignancies: implications for NTRK-targeted therapeutics. *JCO Precis Oncol.* 2018;8:1-20.
 99. Hempel D, Wieland T, Solfrank B, Grossmann V, Steinhard J, Frick A, et al. Antitumor activity of larotrectinib in esophageal carcinoma with NTRK gene amplification. *Oncologist.* 2020;25:e881-6.
 100. Penault-Llorca F, Rudzinski ER, Sepulveda AR. Testing algorithm for identification of patients with TRK fusion cancer. *J Clin Pathol.* 2019;72:460-7.