Fever in a Patient with a Central Venous Catheter Colonized by Pandoraea pnomenusaa

Febre numa Doente com Cateter Venoso Central Colonizado por Pandoraea pnomenusaa

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ABSTRACT
Pandoraea species are a newly described genus of multidrug-resistant, non-fermentative Gram-negative bacilli, mainly isolated from sputum samples of cystic fibrosis patients. In immunocompromised patients or with high antibiotic selective pressure, these pathogens are generally opportunistic and invasive. Although Pandoraea spp. are rare, the true incidence of these infections may be underestimated due to difficulties in microbial identification by phenotypic methods. We present the case of a 51-year-old woman, with new-onset fever after a prolonged hospitalization and multiple courses of antibiotics. Mass spectrometry assays identified Pandoraea pnomenusa in the blood cultures taken from the central venous catheter and in the catheter tip. Fever cessation after catheter removal suggests a catheter-related bloodstream infection. To the best of our knowledge, this is the first isolation of a Pandoraea spp. in Portugal, which should raise awareness to the emergence of these opportunistic and multidrug-resistant microorganisms, and the importance of its prompt identification.

Keywords: Catheter-Related Infections; Central Venous Catheters/adverse effects; Fever; Pandoraea pnomenusa

INTRODUCTION
The Pandoraea genus was first described by Coenye T et al in 2000, after a careful genotypic and phenotypic characterization of species isolated from the environment or sputum of cystic fibrosis (CF) patients, previously misidentified as Burkholderia cepacia, Ralstonia picketti or Ralstonia paucula. The term Pandoraea refers to Pandora’s box in Greek mythology – the origin of the evils of mankind.

These non-fermentative, Gram-negative bacilli (NFGNB), with oxidase activity, are naturally encountered in soil or water, but they can also be nosocomial pathogens associated with invasive devices, such as catheters or ventilation systems.1–6 To date, six species have been identified in humans – Pandoraea apista, Pandoraea pulmonicola, Pandoraea pnomenusa, Pandoraea sputorum, Pandoraea norimbergenis and Pandoraea fibrosa.1,6

Although rare, the Pandoraea spp. are emerging, with most isolates originating from sputum samples of patients with CF or other lung diseases.2,4–7,8 In addition to respiratory samples, isolation in skin lesions, urine and blood have been described in CF and non-CF patients, the latter highlighting the invasive capacity of this species.2,4,9,10

P. pnomenusa was the most isolated species from blood cultures (BC) in a Centers for Disease Control and Prevention study of 2001,2 which revealed an increased potential for invasive disease. Since then, multiple invasive infections of Pandoraea spp. were described, mostly in non-CF patients.5,8–11 Catheter-related bloodstream infections (CR-BSI) and colonization of catheters without bacteremia were also reported.3,12

Phenotypically, Pandoraea spp. are very similar to Burkholderia and Ralstonia genera (with an intermediate phylogenetic position between these two), which may lead to an erroneous phenotypic identification of these agents.1–5,8,10 In fact, the incidence of Pandoraea infections may be underestimated due to difficulties in its identification. However, mass spectrometry assays have proven to be able to identify NFGNB which were not correctly identified by phenotypic methods.5,10,13,14


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The treatment of *Pandoraea* infections can be challenging, as they have a broad resistance to antibiotics, and natural or acquired resistance mechanisms are not yet fully understood.\(^2\)\(^5\)\(^7\)\(^8\)\(^9\). In the particular case of carbapenems, there is a unique discrepancy in terms of susceptibility between meropenem and imipenem. Despite the presence of meropenem-hydrolyzing β-lactamases, which causes intrinsic resistance to meropenem, susceptibility to imipenem is common.\(^2\)\(^4\)\(^5\)\(^8\)\(^9\)\(^10\)\(^12\)

Moreover, due to its rarity, neither the European Committee on Antimicrobial Susceptibility Testing (EUCAST) nor the Clinical and Laboratory Standards Institute (CLSI) have established the antibiotic concentrations (breakpoints) at which this bacteria is considered to be susceptible to a certain antibiotic.\(^5\)\(^9\)

**CASE REPORT**

We report the case of a 51-year-old woman with a past medical history of severe hearing loss of unknown etiology, type 2 diabetes *mellitus* (HbA1c levels between 7.4% and 8.5% before admission), and a recent ischemic stroke requiring hospitalization for 34 days.

Twenty days after her discharge, the patient was admitted to the intermediate care unit with the diagnosis of sepsis due to *Clostridium perfringens* colitis with bacteriemia, and initially completed 14 days of piperacillin/tazobactam and metronidazole.

Due to difficult peripheral access, a central venous catheter (CVC) was placed the following day.

The patient required a total of 78 days of hospitalization, reflecting a complex clinical course, with multiple infections and antimicrobial courses, which are summarized in Fig. 1.

On the 50\(^{th}\) hospitalization day, while on daptomycin and ceftazidime/avibactam, the patient redeveloped subfebrile temperatures (37.4°C) of uncertain origin. BC were drawn the following day. Due to difficulty in obtaining peripheral access, BC were taken from the CVC.

One BC set was positive 21 hours after incubation. On microscopic examination, no bacteria were visualized in Gram staining. On bacteriological cultures, MacConkey agar revealed very small, non-fermentative colonies, oxidase positive, and blood and chocolate agar small, grey and pleomorphic colonies.

On the 53\(^{rd}\) day, due to persistence of fever (maximum of 38.6 ºC) and the presence of a NFGNB on BC, empiric meropenem was initiated. The CVC was removed, and the catheter tip was cultured using a semiquantitative roll plate method (Maki’s technique).

The second BC set was positive after 35 hours of incubation, and the Gram staining revealed very short and thin Gram-negative bacilli.

In both BC taken from the CVC and in the catheter tip culture, *P. pnomenusa* was identified by mass spectrometry (VITEK\(^{2\text{®}}\)MS) with 99.9% certainty. The phenotypic identification method (VITEK\(^{2\text{®}}\)) was only able to identify the agent as *Pandoraea* spp. with 89% certainty.

On the 54\(^{th}\) day, it was possible to obtain two peripheral BC, which yielded negative results.

According to the PK-PD (non-species related breakpoints) of EUCAST 2021,\(^10\) antimicrobial susceptibility was assessed by gradient tests (ETEST\(^{®}\)) for ciprofloxacin, gentamycin, ceftazidime, ceftazidime/avibactam, piperacillin/tazobactam, imipenem and meropenem. The minimal inhibitory concentration of 2 µg/mL of imipenem suggested that the isolated *P. pnomenusa* could only be treated with this antibiotic. For this reason, meropenem was discontinued.

Fever cessation coincided with CVC removal and no further treatment was instituted. The patient remained apyretic until discharge.

**DISCUSSION**

We report the identification of a rare microorganism – *P. pnomenusa*. To the best of our knowledge, this is the first isolation of *Pandoraea* spp. in Portugal.

The correct identification of NFGNB is crucial as the true incidence of *Pandoraea* infections may be underestimated. Its identification should be performed with mass spectrometry assays, which have demonstrated to have greater diagnostic accuracy compared with other phenotypic methods. In our laboratory VITEK\(^{2\text{®}}\) was not able to distinguish between *Pandoraea* species.

It is known that *Pandoraea* spp. are opportunistic pathogens, infecting mainly CF patients, or individuals with some degree of immunosuppression or with a high antibiotic selective pressure.\(^3\)\(^5\)\(^8\)\(^9\)\(^10\)\(^12\) Our patient was hospitalized for a long period of time and received multiple antibiotic therapies, which probably favored the appearance of this organism.

Although *P. pnomenusa* was isolated in the BC taken from the CVC and in the catheter tip, the diagnosis of CR-BSI cannot be confirmed, as this agent was not identified in simultaneous peripheral BC. On the other hand, the fever had no-localized symptoms and ceased after removal of the colonized CVC, which supports this diagnosis.

To conclude, clinicians should be aware of the emergence of these multidrug-resistant microorganisms in susceptible patients, as they can be both colonizers and invasive infectious agents. Since *Pandoraea* species are a natural reservoir of carbapenem-hydrolyzing oxacillinases (antibiotic resistance genes), empirical treatment with carbapenems may be inappropriate, and even result in increased morbidity. Furthermore, the physical proximity with other species may promote...
plasmid-mediated horizontal gene transfer of these resistance genes, favoring the appearance of new multi-drug resistance pathogens.

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AUTHORS CONTRIBUTION
SRO: Literature review, data acquisition, drafting of the paper.
ICM: Literature review, data acquisition.
CR: Critical review of the paper (medical data).
MJS: Critical review of the paper (laboratory data).

PROTECTION OF HUMANS AND ANIMALS
The authors declare that the procedures were followed according to the regulations established by the Clinical Research and Ethics Committee and to the Helsinki Declaration of the World Medical Association updated in 2013.

DATA CONFIDENTIALITY
The authors declare having followed the protocols in use at their working center regarding patients’ data publication.

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**Figure 1** – Infections and antibiotics during hospital stay

- Admission
- PN initiated
- CVC implanted
- CVC switched
- CVC removed
- New-onset fever
- CVC re-implanted
- CVC removed and PN suspended

**Infants**

- Colitis by *Clostridium perfringens*, with bacteremia
- Megacolon by Citomegalovirus
- Candidemia by *Candida parapsilosis*
- Bacteremia by methicillin-resistant *Staphylococcus epidermidis*
- Low urinary tract infection by *carbapenemase-producing Klebsiella pneumoniea*
- No infectious agent was found

**Antibiotics**

- Piperacillin/tazobactam + metronidazole
- Ganciclovir/vaganciclovir
- Micafungin
- Vancomycin
- 3 doses of fosfomycin
- Daptomycin
- Ceftazidime/avibactam

**Timeline**

- Time in days of hospital stay:
- F: Infection and infectious agent; CVC: central venous catheter; PN: parenteral nutrition.

*On the 39th day, the patient initiated fever without localized symptoms. No infectious agent was found, so daptomycin and ceftazidime/avibactam were initiated empirically.*